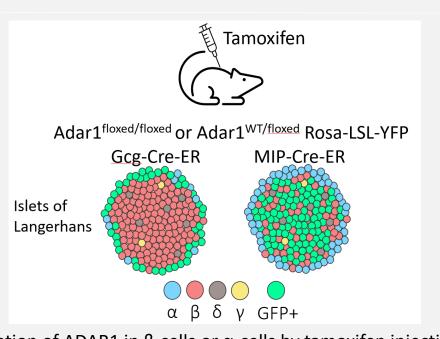
Institute for Medical Research Research Strate-Canada The role of RNA editing in α and β cells with research Resear

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Background

RNA editing, involving adenosine deamination by ADAR1, serves to dismantle endogenous double-stranded RNA (dsRNA) structures that could potentially mimic viral infection and trigger autoinflammation. Despite its emerging central role as a regulator of inflammation, A-to-I RNA editing has not been studied in pancreatic islets.

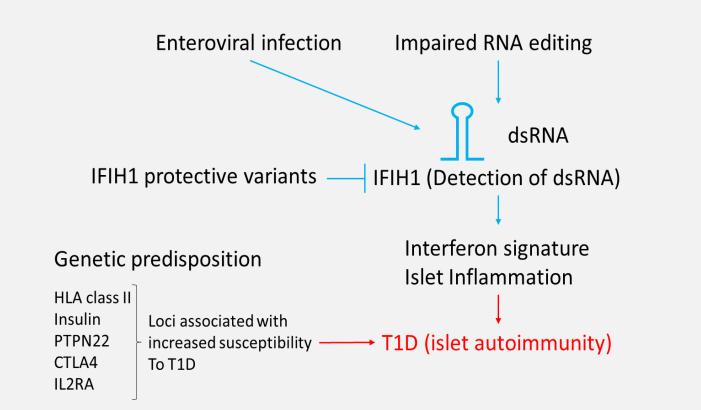
We hypothesize that RNA editing serves to limit autoinflammation in islets, and that its disruption may model early stages of type 1 diabetes, involving an interferon response. To test this hypothesis we characterized the impact of disrupted RNA editing in mouse islets, using tamoxifen-inducible deletion of ADAR1 in β -cells or α -cells in vivo.



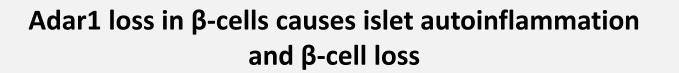
1. Method : mouse models

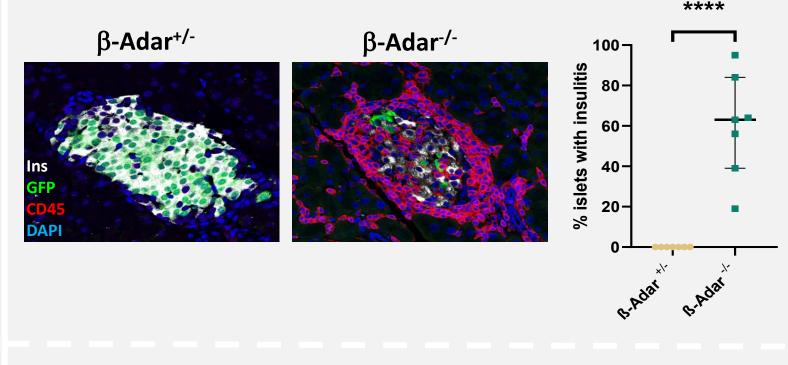
Deletion of ADAR1 in β-cells or α-cells by tamoxifen injection to 1 month-old Insulin-CreERT or Glucagon-CreERT; Adar1 F/F; Rosa-LSL-YFP mice. This results in disruption of RNA editing in adult β-cells/α-cells.

2. Model for possible involvement of impaired RNA editing in the etiology of T1D



3. Effects of <u> β -cell-specific</u> ADAR1 knockout

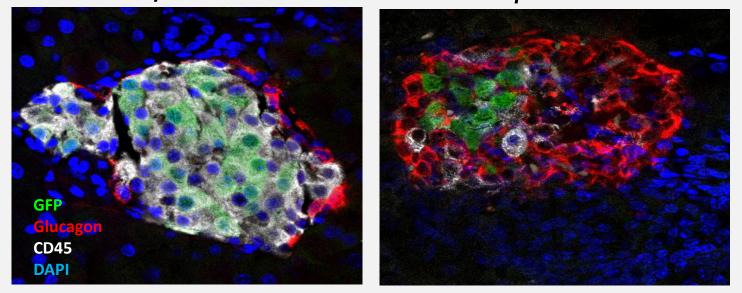




4. Preservation of α-cells in β-cell-specific ADAR1 knockout

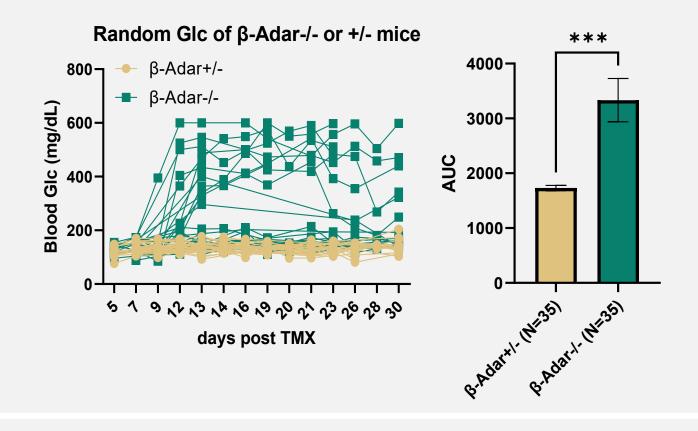
β-Adar^{+/-}

β-Adar^{-/-}

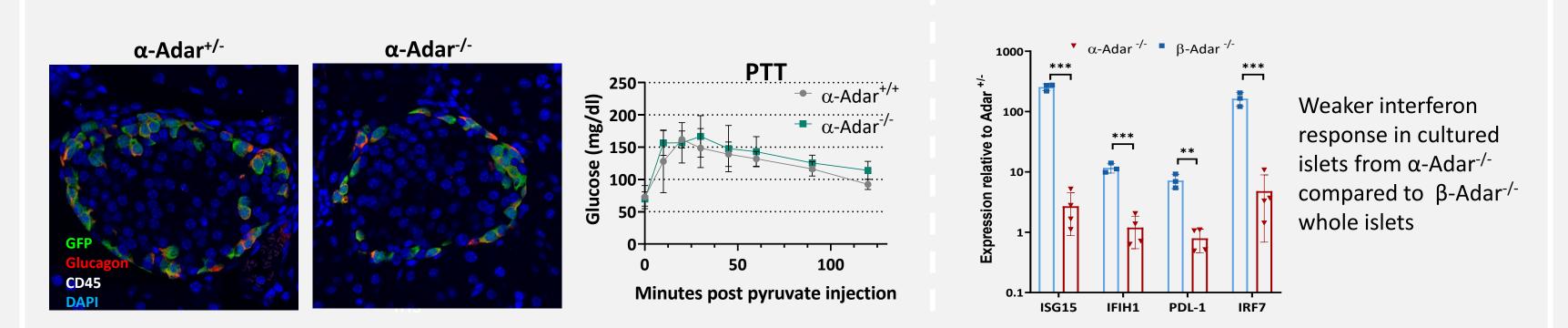


Results so far suggest that α -cell numbers remain unchanged in β -Adar^{-/-} mice, despite massive islet inflammation and beta cell destruction.

Adar1 loss loss in β-cells causes disruption of glucose homeostasis



5. <u> α -cell-specific</u> ADAR1 knockout does not elicit inflammation and does not impair α -cell function



Conclusions

Disruption of ADAR1 specifically in β-cells causes massive islet inflammation, β-cell destruction and diabetes.
Despite extensive immune response in islets, α-cells remain unscathed, reminiscent of α-cell persistence in T1D.
Neither an immune response nor a metabolic phenotype is observed upon ADAR1 disruption in α-cells of young mice.
Cultured islets from α-Adar^{-/-} mice elicit a much weaker interferon response compared with islets from β-Adar^{-/-} mice, suggesting an intrinsic resistance of α-cells to disrupted RNA editing.
Ongoing experiments are aimed at understanding the molecular basis for the unique resistance of α-cells to ADAR1 deficiency.